# AMMA JOURNAL VOL 13 ISSUE 2 OCTOBER 2004 BIOLOGICAL AGENTS Genetic Vaccination: Can Plasmid DNA deliver its expectations? <sup>1</sup>

by Malcolm R. Alderton, Peter J. Gray and David F. Proll<sup>2</sup>

VACCINES ARE EFFECTIVE AT protecting not only individuals but also communities. In the last fifty years, global immunisation programs have developed such strong "herd immunity" that some diseases, e.g. smallpox and polio, have been effectively eliminated.

Currently, in terms of commercially available vaccines, the traditional methods of vaccine development are still leading the attack against infectious agents. These vaccines are produced from dead or an attenuated (non-pathogenic) form of the pathogen, which induces immunity in an individual and protects against clinical disease. The first of these- attenuated vaccines - cause a mild infection using a strain or serotype of the pathogen that has reduced pathogenicity.

Examples of this type of vaccine include the original smallpox vaccine pioneered by Jenner, and those currently used to protect against polio (Sabin), tuberculosis (BCG), typhoid, measles, mumps, yellow fever and rubella. However, live attenuated vaccines carry the risk, albeit very low, that the live attenuated preparations may revert to pathogenic forms. To circumvent the risks associated with live attenuated vaccines, killed microbes, toxoids and recombinant subunit preparations have been developed to protect against diseases such as the plague, Q fever, hepatitis A and B, tetanus and diphtheria. These vaccines are safer than live attenuated vaccines but do not stimulate the same level of immunity.

Because of a lack of interest by commercial vaccine producers in the diseases caused by potential biological warfare (BW) agents (Table 1), the vaccines currently available are first-generation vaccines. These often require a course of vaccinations with regular boosters and are therefore not ideal for military applications. The ideal vaccine for military use should possess the following characteristics:

- i) induce rapid and complete protection,
- ii) require only one oral immunization to reduced logistics burden and cost,
- iii) safe with no debilitating side effects,
- iv) provide life-long immunity without the requirement of boosters vaccinations,
- v) stable at high temperatures thereby eliminating the need of refrigeration and increasing the shelf-life of the vaccine, and
- vi) low production costs

# NEW METHODS IN VACCINE DEVELOPMENT

Although there have been successes with traditional vaccination, current vaccines target only a tiny fraction of infectious diseases. Table 1 lists possible biological warfare agents and highlights the fact that of the 23 organisms/toxins listed, only 11 currently have vaccines available. Furthermore, the increasing vulnerability of human populations to new and re-emerging infections, and the dramatic rise in antibiotic-resistant micro-organisms, has indicated the urgent need for new vaccine research. Vaccines are an effective way of preventing disease but traditional vaccine strategies, based on either

an attenuated or inactivated micro- organism, have provided limited or no protection against some pathogens. For example, tens of millions of dollars have been spent over the last 30 years to develop a traditional vaccine to prevent malaria, without success. However, advances in biotechnology and genetic engineering techniques have enabled new approaches to vaccine development - such as DNA vaccines.

## **DNA VACCINES**

It has been a decade since the discovery that the injection of naked DNA into muscle cells, resulted in long-term gene expression in transfected cells. Subsequently, it was shown that if the plasmidencoded an antigenic protein, it was possible to elicit an immune response. From this spawned a new era in vaccine development, and hence the technology of DNA (or genetic, nucleic acid) vaccination. The application and design of novel DNA vaccines grew rapidly. From the beginnings in 1990-1992, it was only three years later that the first human trials with a DNA vaccine began <sup>5</sup>. Despite this, to date, not a single DNA vaccine has been licensed for human use. It may be asked, therefore, if there is a future for this technology. The mechanisms underlying DNA vaccination are discussed below, out-lining the current developments and future prospects and its application to the development of vaccines for military application.

AGENT	DISEASE	VACCINE
Bacteria		
Bacillus anthracis	Anthrax	Yes
Brucella spp.	Brucellosis	No
Vibrio cholerae	Cholera	Yes
Burkholderia mallei	Glanders	No
Burkholderia pseudo mallei	Melioidosis	No
Yersinia pestis	Plague	Yes
Francisella tularensis	Tularemia	Yes, but investigational
Rickettsia		
Coxiella Burnetii	Q Fever	Yes, Australia only
Rickettsia prowazekii	Typhus	No
Viruses		
Variola virus	Smallpox	Yes
Venezuelanequine	VEE	Yes
encephalitis (VEE) virus		
Ebola virus	Viral hemorrhagic fever	No
Marburg virus	Viral hemorrhagic fever	No
Argentine hemorrhagic fever virus	Viral hemorrhagic fever	Yes, but investigational
Congo-Crimean hemmorhagic	Viral hemorrhagic fever	Νο
fever virus	Viral hemornagic level	
Rift valley fever virus	Viral hemorrhagic fever	Yes, but investigational
Hantvirus	Viral hemorrhagic fever	No
Yellow fever virus	Viral hemorrhagic fever	Yes
Dengue fever virus	Viral hemorrhagic fever	No
Toxins		
Botulinum toxin	Botulism	Yes
Ricin	Ricin poisoning	No

Staphylococcal enterotoxin B	SEB intoxication	No
(SEB)		

## Table I. Availability of vaccines to potential biological warfare agents. <sup>5</sup>

DNA vaccines are considered to be the technology that will address the efficacy problems of the current vaccines and are often referred to as the third generation of vaccines. The methodology was an offshoot of gene therapy, a technology developed to deliver genes coding for proteins that could replace a defective enzyme or tag a cancer cell for destruction.

In the search for an appropriate vehicle that would deliver the DNA to cells in a way that would promote uptake and expression of the DNA, it was discovered that injection of DNA plasmids in saline stimulated expression of foreign proteins in mice. Unfortunately, the amounts of protein produced by the tissue receiving the plasmids were not enough to correct an enzyme defect or provoke the destruction of a tumour, but the levels were high enough to stimulate an immune response.

In its simplest form, a DNA vaccine consists of the DNA sequence encoding the potential antigen under the control of a eukaryotic promoter to drive gene expression in the host cell. However, they generally consist of a bacterial plasmid engineered for gene expression in eukaryotic cells. Thus, they contain a eukaryotic promoter and a transcription termination/ polyadenylation signal sequence to aid in mRNA stability. In association with this, they also have a bacterial origin of replication and an antibiotic selectable marker to enable growth and manipulation in bacterial cells (Figure I).

## Antibiotic

Promoter Plasmid backbone – signals and genes for growth/manipulation in bacteria Transcription unit – for antigen synthesis

Figure 1: Schematic representation of a DNA vaccine. Plasmid DNA used for vaccination is composed of two domains. A transcription unit composed of a promoter to drive mRNA synthesis and polyadenylation I termination signal to correctly process the transcribed mRNA. These two regions flank the sequence coding for the antigen to be **synthesised: The second** domain enables the plasmid to be genetically manipulated in bacterial cells during the production and growth of the plasmid DNA vaccine.

The antigen-encoding DNA molecule has been delivered into the host by a wide variety of methods and routes. These include: (i) into the skin via the epidermal gene gun, intradermal or subcutaneous injection, (ii) into the nasal and gastrointestinal mucosa via intranasal and oral delivery, (iii) into the bloodstream by intravenous injection, and (iv) into the genitourinary tract by intravaginal injection or installation. <sup>10</sup> Following inoculation of the plasmid DNA into the host, transcription/translation occurs to produce a mature protein (Figure 2). The protein is then exposed to the host's immune cells resulting in an immune response.

A variety of immune responses have been characterised following vaccination with DNA molecules. Generally, the response by DNA immunisation is characterised as a T helper cell type 1 (Th 1) response, rather than a T helper type 2 (Th2) response.<sup>11</sup> Th1 are inflammatory cells that are associated with the cell-mediated response, whereas, Th 2 are associated with antibody production.

FIGURE 2: Schematic representation for the production of antigens from DNA vaccines within a host cell. Following the introduction of the plasmid DNA into the cell (I), mRNA is produced in the nucleus (NU), through transcription from the gene encoding the antigen (2). Export of the mRNA molecules from the nucleus into the cytoplasm (3) allows interaction with the cells translational apparatus to produce mature proteins (4). The antigens that are engineered to contain secretion signals or signals to target them to immune cells are secreted from the cell (5) and interact or travel to immune cells for processing. Alternatively, the antigens that do not contain secretion signals are proteolytically degraded into smaller peptides (6) and transported to the endoplasmic reticulum (ER). Within this cellular organelle, the peptides become associated with the Major Histocompatibility Complex (MHC) (7) and transported to the outer cell membrane where the MHC associated antigen is exposed on the outer cell surface for interaction with immune cells.

However, this characterisation is not all or nothing choice between Th1 and Th2 immunity but rather a dominant response profile that depends on a series of factors, including the antigen, the dose, the route and method of delivery, whether the antigen is secreted, and if adjuvants or immunostimulators are used<sup>12.15</sup>. On the other hand, traditional vaccines mostly, and sometimes exclusively, generate a humoral response. As a result DNA vaccines are particularly well suited to deal with viral diseases against which traditional vaccines have failed, or have not been produced.

## **ADVANTAGES OF DNA VACCINES**

DNA vaccines have several advantages over traditional vaccines. Studies have shown them to be well tolerated, and stimulatory of a full spectrum of immune responses, including cytotoxic T lymphocytes (CTL), a response generally not induced by protein-based vaccines. The induction of CTL, by virtue of the in vivo antigen synthesis and MHC class I molecule presentation, provides the efficacy of a live attenuated vaccine without the risk of infection. They also generate exceptionally long-lasting immune responses, <sup>16.18</sup> thus possibly reducing the need for booster immunisations.

There is also the possibility of producing multivalent vaccines against several pathogens by cloning genes encoding for different antigens into a single vector, or by mixing different plasmids together. The ease of cloning allows new vaccines to be created quickly, for example in response to new or changing strains of pathogens.

Furthermore, the new generation vaccines will be more cost-effective, since they can all be produced using similar techniques. The simplicity of producing DNA vaccines is one of the most attractive aspects of the process; the only steps required being the construction and purification of the plasmid. The robustness of DNA eliminates the need for cold storage, so the cost of storage and transport will also be reduced. These characteristics make DNA vaccinology very attractive to the military, which desires greater disease protection for combat personnel, with fewer immunisations, and at less cost.

## **DNA VACCINE SAFETY ISSUES**

Although the immunogenicity of DNA vaccines is well established, there are still some concerns regarding their safety, including the integration of DNA plasmids into the host chromosome, the production of antibodies to DNA and the generation of neonatal antigen tolerance<sup>1</sup>. Integration of vaccine DNA into the chromosome is perceived as a problem because of the possible activation of oncogenes or the disruption of normal gene function and regulation. Clearly, these events could have serious consequences for the host, but in studies, to date, the integration of plasmid DNA into the host

chromosome has not been reported. Shroff and colleagues<sup>10</sup> were unable to detect plasmid integration in several preclinical studies using an assay that can detect one integration event in 1 x 106 cells. Davis and McCiuskie<sup>15</sup> argue the likelihood of integration of the DNA vaccine plasmid into the host chromosome is very low, since, (i) most injected DNA is rapidly degraded in the extracellular space, (ii) the plasmid DNA vectors are designed to remain episomal, (iii) most non-integrated DNA would soon be lost during subsequent cell division, and (iv) integration is not possible in mature muscle fibres, as they are permanently post-mitotic.

The production of anti-DNA antibodies is also of concern, especially to individuals with autoimmune diseases, e.g. systemic lupus erythematosus (SLE). Although double-stranded DNA can induce low levels of antibodies<sup>11</sup> the fear that this will initiate unwanted autoimmune reactions has not materialised. In addition, Mor *et al.*<sup>21</sup> also suggested that neonatal immunisation with a DNA vaccine plasmid generated immune tolerance but all subsequent studies using newborn animal hosts have not substantiated this claim. Although significantly more work is required on the safety of DNA vaccines, the work to date suggests most of the concerns are likely to prove unfounded.

## HUMAN CLINICAL TRIALS

Despite the wealth of accumulated data on the vaccination of animals with DNA and the subsequent characterisation of the immune response, until recently their effectiveness in humans was unknown. Due in part to the safety concerns discussed above, the rapid advancement of the technology for application to human disease had stalled somewhat. However, with investigations now addressing these concerns, human trials have slowly begun -albeit with caution and scepticism.

One of the first human trials of DNA vaccination was against the malaria 1 parasite Plasmodium falciparum,<sup>12</sup> These phase 1 clinical trials established the safety, tolerability and immunogenicity of DNA vaccines encoding malarial antigens. They also established their capacity to induce antigen-specific CDS+T cell-dependant CTL and I NF- responses in humans by three different routes of administration. However, antibodies were not produced despite the fact that the vaccines used had previously been shown to induce antibody responses in a range of animals including primates.

Similar phase 1 clinical trial have also evaluated HIV and Hepatitis B DNA vaccines. Not only were a number of safety issues monitored, but also the ability of the DNA vaccines to induce an immune response measured. A range of parameters was examined, including the amount and isotype of antibody produced against the encoded proteins, the production of cytokines, the activation and proliferation of peripheral blood mononuclear cells and for antigen specific cytotoxic activity.<sup>3 25</sup> Although both B and T cell immune response has been reported following the phase 1 trials, the magnitude of the elicited responses were modest.

Although the results obtained from these preliminary trials were somewhat mixed they did provide evidence that the concept of DNA vaccination works in humans, and generated some invaluable data in relation to the dose and route of administration suitable for humans. The results also demonstrated that the observed responses were suboptimal for effective vaccines and needed to be significantly improved to yield increased immune responses. This has now paved the way for future trials to address these problems.

#### FUTURE PROSPECTS AND DEVELOPMENTS

Despite the early optimism for the potential of DNA vaccines, much of the work to date has found that the immune responses induced through injection of DNA are insufficient to provide immunity against

subsequent challenge by the infectious agent. Thus, more recently much effort has focused on means to increase and direct more specifically the immune response by modifying the plasmids and/or their mode of delivery.

## **Co-delivery of cytokines**

Cytokines and co-stimulatory cell surface molecules play a crucial role in directing and determining the the magnitude of an immune response. Consequently, numerous workers have used plasmid DNA encoding various cytokines and co-stimulatory molecules to enhance or direct the immune response generated following DNA vaccination.<sup>10</sup> These included the cytokines: Interleukins1, 2, 4, 5, 6, 7, 8, 10, 12, 15 and 18, Granulocyte-macrophage colony-stimulating factor (GM-CSF), Tumour necrosis factor (TNF) and Interferon -g. Generally, these have resulted in increased antibody responses while some have also increased T-cell responses.

## **Delivery mechanisms**

Clearly, there is a wide variety of ways of administering DNA vaccines and each may have advantages specific to the vaccine in question. There is, however, a problem of DNA degradation in the vaccine recipient. The extent of the DNA degradation by extracellular deoxyribonucleases may vary for the different routes of administration, but approaches to protect the DNA vaccine and assist efficient entry into host cells are imperative in DNA vaccine design.<sup>20</sup>

Sizemore *et al* <sup>28</sup> used a highly attenuated Shigella spp to deliver and express DNA vaccines in mucosal surfaces. The bacterium, which enters the mucosa via M cells, evades phagosome formation in phagocytic cells by entering the cytosol. This improves vaccine efficacy by initiating a strong CTL response as well as a humoral response. Other microorganisms that generate strong cell-mediated responses and could be used in a similar way to deliver antigens encoded by DNA vaccines include attenuated Salmonella typhimurium, Listeria monocytogenes, Leishmania major and Mycobacterium tuberculosis.

Gregoriadis *et al* <sup>29</sup> identified uptake and expression of DNA vaccines by antigen-presenting cells (APC), in particular dendritic cells, which is preferable to uptake and expression by muscle cells. In 1990, he also demonstrated liposomes were avidly taken up by APC, and later revealed liposome coated DNA vaccines were also preferentially targeted by APC. The liposomes also protect the DNA vaccines from deoxyribonuclease degradation. Moreover, a number of liposome-based drug formulations, including a hepatitis A vaccine, have been license in the USA and Europe, making future licensing of liposome coated DNA vaccines more likely.

Cochleates are rigid, calcium-induced structures consisting of spiral bilayers of anionic phospholipids<sup>30</sup> a unique structure that differs from liposomes. It is believed that following fusion of the cochleate with the cell membrane the contents (i.e. the DNA vaccine) are released into the cytosol DNA vaccines delivered encapsulated in cochleate have been shown to promote strong long-lasting humoral, mucosal and cell-mediated responses<sup>31</sup>.

Another potential delivery method for DNA vaccines is the use of biodegradable microparticles. DNA contained within microparticles, composed of polylactide-co-glycoside, can be given systemically or to mucosal surfaces. The ability of microparticle encapsulated DNA vaccines to induce mucosal and systemic immune responses has been demonstrated<sup>31-33</sup>. Thus by protecting the DNA vaccine from extracellular elements a more enhanced immune response can be generated, which is probably due to increased antigen production.

## **Targeting antigen molecules**

Incorporating signalling sequences within the vaccine antigen that directs it to the cytosol or endosome in antigen-presenting cells, eg macrophages and dendritic cells ensures the antigen is associated with MHC Class I and Class II molecules, respectively. Antigens associated with MHC Class I molecules initiate a cell-mediated immune response, whereas antigens associated with MHC Class II molecules initiate a humoral response.<sup>3435</sup> The ability to direct antigens to the most appropriate cell and/or cellular compartment would greatly benefit vaccine production, by generating the most appropriate immune response for the particular disease.

Addressins are surface molecules expressed by several different cells including subpopulations of lymphocytes and are responsible for directing these cells to the appropriate lymphoid tissue. They interact with specific receptors on cells located in these tissues, eg thymus, spleen, lymph node, Peyer's patch and bone.<sup>36</sup> Fusing vaccine antigens to these homing molecules may provide a way of transporting the antigens to sites where the pathogen will be first encountered. Experiments performed by Boyle el at.,<sup>37</sup> using DNA vaccines encoding antigen-ligand fusions targeting the antigen to lymphoid organs, showed a significant increase in both the humoral and the cellular immune response.

## **Expression Library Immunisation**

Despite the progress in the development of DNA vaccines, one of the greatest hurdles that still remain in the identification of suitable antigens that will provide protection against disease. An approach to circumvent this may be through the use of Expression Library Immunisation (ELI). The ELI approach was first described by Barry et al<sup>38</sup> who applied the technique to protect mice against Mycoplasma pulmonis. A similar approach has also been used to protect mice against other parasites, including malaria (personal observations)<sup>39</sup> and Leishmania<sup>40</sup>. In this approach, the entire genome (in small fragments) is cloned into a eukaryotic expression vector similar to those used in DNA vaccination, and then the entire library (or fractions thereof) is used to vaccinate the subjects. Using a mathematical model, sufficiently large enough numbers of plasmids are generated to ensure the probability that every gene contained in the organism's genome is represented in the library. For example, the mouse infecting malaria parasite, Plasmodium chabaudi, which possess a genome of approximately 25-30Mb, requires approximately 70,000 plasmids (each containing a median size of 1.5kb of P. chabaudi DNA) in order for the entire genome to be represented. However, not all genes encode "protective antigens" therefore a significant reduction in the number of plasmids may still provide protective immunity. This is highlighted using the example above where protection against a lethal challenge of malaria was observed when mice are vaccinated with a subset of the entire library containing as little as 3,000 plasmids (personal observations).<sup>39</sup> Further sequential partitioning of these subsets could lead to the identification of individual protective antigens - a technique that may have generic applicability to the discovery of vaccine antigens.

ELI has enormous potential for military application. It is feasible that the concept could be applied to the battlefield. A potential vaccine, which could provide suitable protection following the onset of disease, could be produced as follows. A soldier falls ills and the causative organism is isolated (2 days). The organism is grown and genomic DNA purified and used to construct an expression library (2-3 days). The library is expanded to produce sufficient amounts of DNA to vaccinate other soldiers (2 days). Hence, within less than ten days from the onset of illness, a vaccine could be produced. The reason for the rapid speed at which this type of vaccine can be produced lies in the fact that because (at least in mathematical theory – determined by the number of individual DNA plasmids) every protein encoded by the target organism is contained within the library. Thus eliminating the need to identify

and characterise specific protective antigens. The US recently announced a large project, to be headed by Stephen L Hoffman of the Navel Medical Research Centre, to investigate further the application of ELI technology to military applications.

# CONCLUSIONS

The development of DNA vaccine technology has been rapid, with the progression from a laboratory phenomenon to clinical trials occurring at breakneck speeds. The DNA vaccine vacuum fuelled a desire to harness the tools of molecular biology to construct antigen-encoding plasmids capable of inducing protective immune responses to pathogens for which there were no conventional vaccines. However, many questions regarding the basic science of DNA vaccine technology remain unanswered - and this will ultimately determine the success or failure of the application to find a place in our immunological arsenal against disease.

# REFERENCES

- 1. M1MS. MIMS Annual 1998.Sydney: Griffin Press; 1998.
- 2. Cleland JL. Single-administration vaccines: Controlled-release technology to mimic repeated immunizations. Trends Biotechnology 1999;17(1):25-9.
- 3. Wolff JA, Malone RW, Williams P, Chong W, Acsadi G, Jani A, Feigner PL. Direct gene transfer into mouse muscle in vivo. Science 1990; 247:1465-8.
- 4. Tang D, De Vit M Johnston SA. Genetic immunization is a simple method for eliciting an immune response. Nature 1992; 356:152-4.
- 5. Martin I, Parker SE, Hedstrom R, Lel Hoffman SL, Norman J, Hobart P, Lew D. Plasmid DNA malaria vaccine: The potential for genomic integration after intramuscular injection. Hum Gene Tiler 1999;10(5):759-68.
- Parker SE, Borellini F, Wenk ML, Hobart P, Hoffman SL, Hedstrom R, LeT, Norman JA. Plasmid DNA malaria vaccine: Tissue distribution and safety studies in mice and rabbits. Hum Gene Tiler 1999;10(5): 741-58.
- Hedstrom RC, Doolan DL, Wang R, Kumar A, Sacci JB Jr., Gardner MJ, Aguiar JC, Charoenvit Y, Sedegah M, Tine JA, Margalith, Hobart P, Hoffman SL. In vitro expression and in vivo immunogenicity of Plasmodium Falciparum pre-erythrocytic stage DNA vaccines. Int j Mol Med 1998;2(1):29-38.
- Gramzi nski RA, Obaldia N III, Jones TR, Rossan RN, Collins WE, Garrett DO, La AA, Hoffman SL. Susceptibility of Panamanian Aotus Lemurinus Lemurinus to sporozoite- induced Plasmodium Falciparum (Santa Lucia) infection. Am) Ti-op Med Hyg 1999;61(1):19-25.
- Wang R, Doolan DL, Le TP, Hedstrom RC, Coonan KM, Charoenvit Y, Jones TR, Hobart P, Margalith M, Ng], Weiss WR, Sedegah M, de Taisne C, Nonnan JA, Hoffman SL. Induction of antigen-specific Cytotoxic I Lymphocytes in humans by a Malaria DNA vaccine. Science 1998;282(5388):476-80.
- 10. Guruna than S, Klinman DM, Seder RA. DNA vaccines: Immunology, application, and optimization. Annual Rev Immunology 2000; 18:927-74.
- Shroff KE. Marcucci LA, de Bruin Sj, Winter LA, Tiberio L, Pachuk C, Synder LA, Satishchandran C, Ciccarelli RB, Higgins Tj. Induction of HSV-GD2 specific CD4(+) cells in Peyer's patches and mucosal antibody responses in mice following DNA immunisation by both parenteral and mucosal administration. Vaccine 1999;18(3-4):222-30.
- 12. Barry MA, Johnston SA. Biological features of genetic immunization. Vaccine 1997; 15:788.
- 13. Feltquate OM, Heaney S, Webster RG, Robinson HL. Different Th cell types and antibody isotypes generated by saline and Gene Gun DNA immunization. J Immunology 1997;158.

- 14. Torres CA, Iwasaki A, Barber BH, Robinson HL. Differential dependence of target site tissue for Gene Gun and intramuscular DNA immunizations. j Immunology 1997; 158:4529.
- 15. Davis HL, McCiuskie MJ. DNA vaccines for viral diseases. Microbes Infect 1999; 1:7-21.
- 16. Yankaukas MA, Morrow JE, Parker SE, Abai A, Rhodes GH, Dwarki Vj, Gromkowski SH. Longterm Anti-Nucleoprotein cellular and humoral immunity is induced by intramuscular injection of Plasmid DNA containing NP Gene. Cellular Biology 1993; 12:7712-7.
- 17. Davis HL, Michel M-L, Whalen RG. DNA-based immunization induces continuous secretion of Hepatitis B surface antigen and high levels of circulating antibody. Human Molecular Genetics 1993; 2:1847-51.
- Lazes E, Huygen K., Content K, Dennis O, Montgomery DL, Yawman AM, Vandenbussche P, Van VoorenJP, Drowart A, Ulmer JB, Lui MA. immunogenicity and efficacy of a Tuberculosis DNA Vaccine encoding the components of the secreted Antigen 85 Complex. Vaccine 1997; 15:830-4.
- 19. Smith H A, Klinman OM. The regulation of DNA vaccines. CurT Opin Biotechnology 2001;12(3):299-303.
- 20. Shroff KE, Smith LR, Baine Y, Higgins Tj. Potential for Plasmid DNAs as vaccines for the new millennium. Pharmaceut Science Technology Today 1999;2(5):205-12.
- 21. Mor G, Yamshchikov G, Sedegah M, Takeno M, Wang R, Houghten RA, Hoffman S, Klinman OM. Induction of neonatal tolerance by plasmid DNA vaccination of mice. j Clin Investigations 1996; 98:2700-5.
- 22. Doolan DL, Hoffman SL. DNA-based vaccines against malaria: Status and promise of the multistage Malaria DNA Vaccine operation. Int j Parasitol 2001;31(8):753-62.
- 23. MacGregor RR, Boyer JD, Ugen KE, Lacy KE, Gluckman Sj, Bagarazzi ML, Chanergoon MA, Baine Y, Higgins Tj, Ciccarelli RB, Coney LR, Ginsberg RS, Weiner DB. First human trial of a DNA-Based vaccine for the treatment of Human Immunodeficiency Virus Type 1 infection: Safety and host response. j Infect Dis 1998;178(1):92-100.
- 24. Tacket CO, Roy Mj, Widera G, Swain WF, BroomeS, Edelman R. Phase 1 safety and immune response studies of a DNA Vaccine encoding Hepatitis B Surface Antigen delivered by a gene delivery device. Vaccine 1999;17(22):2826-9.
- 25. Calarota S, Bran G, Nordlund S, Hinkula j, Lea Andersson AC, Sandstrom E, Wahren B. Cellular cytotoxic response induced by DNA Vaccination in HI V-1- infected patients. Lancet 1998;351(9112):1320-5.
- 26. Manicka n E, Karem KL, Rouse BT. DNA Vaccines- A modern gimmick or a boon to Vaccinology. Critical Rev Immunology 1997.17(2):139-54.
- 27. Gregorian is G. Genetic Vaccines: Strategies for optimisation. Phannaceut Research 1998;15(5):661-70.
- 28. Sizemore DR, Branstrom A, Sadoff]. Attenuated Shigella as a DNA delivery vehicle for DNAmediated immunogenicity. Science 1995; 270:299-302.
- 29. Gregoriadis G, Saffie R, DeSouzajB. Liposome-mediated DNA vaccination. FEBS Letters 1997; 402:107.
- 30. Papahadjopoulos D, Vail Wj, Jacobson K, Poste G. Cochleate lipid cylinders: Formation by fusion of unilamellar vesicles. Biochemical Biophysica Acta 1975; 394:483-91.
- 31. Gou ld-Fogerite S, Mannino Rj. Mucosal and systemic immunization using Cochleate and Liposome Vaccines. j Liposome Research 1996; 6:357-79.
- 32. Mannino RJ, Gould-Fogerite S. Lipid matrix-based vaccines for mucosal and systemic immunization. Pharm Biotecl111ol 1995; 6:363-87.
- 33. Gould-Fogerite S, Kheiri MT, Za ng F, Wang Z, Scolpino AJ, Feketeova E, Canki M. Mannino Rj. Targeting immune response induction with Cochleate and Liposome-based vaccines (Review). Adv Drug Delive1y Rev 1998;32(3):273-87.

- 34. Isobe H, Morant, Li S, Young A, Nathenson S, Palese P, Bona C. Presentation of a Major Histocompatibility Complex Class I Molecule of Nucleo- Protein Peptide expressed in two different genes of an Influenza Virus Transfectanl. j Experiment Med 1995; 181:203-13.
- 35. Schutze-Redelmeier MP, Gournier H, Garcia-Pons F, Moussa M, Joliot A, Volovitch M, Prochiantz A, Lemmonier FA. Introduction of endogenous antigens into the MHC Class I processing and presentation pathway by Drosophila Antennapedia Homeodomain primes Cytotoxic T Cells in vivo. j Immunology 1996; 157:650-5.
- 36. Kraal G, Mebius RE. High endothelial venules: Lymphocyte traffic control and controlled traffic. Advances Immunology 1997; 65:347-95.
- 37. Boyle JS, Brady JL, Lew AM. Enhanced responses to a DNA vaccine encoding a fusion antigen that is directed to sites of immune induction. Nature 1998;392(6674):408-11.
- 38. Barry MA, Lai WC, Johnston SA. Protection against Mycoplasma infection using Expression-Library Immunization. Nature 1995;377(6550):632-5.
- 39. Smooker PM, Setiady YY, Rainczuk A, Spithill TW Expression Library Immunization protects mice against a challenge with virulent rodent malaria. Vaccine 2000;18(23):2533-40.
- 40. Piedrafita D, Xu D, Hunter D, Harrison RA, Liew FY. Protective immune responses induced by vaccination with an expression genomic library of Leishmania Major. Immunol 1999;163(3):1467-72.
- 41. Geissler E. Biological and Toxin Weapons Today. Oxford: Oxford University Press; 1986.